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Spatiotemporal changes of pelagic food webs investigated by environmental DNA metabarcoding and connectivity analysis

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Environmental DNA metabarcoding (eDNA metaB) is fundamental for monitoring marine biodiversity and its spread in coastal ecosystems. We applied eDNA metaB to seawater samples to investigate the spatiotemporal variability of plankton and small pelagic fish, comparing sites with different environmental conditions across a coast-to-offshore gradient at river mouths along the Campania coast (Italy) over 2 years (2020–2021). We found a marked seasonality in the planktonic community at the regional scale, likely owing to the hydrodynamic connection among sampling sites, which was derived from numerical simulations. Nonetheless, spatial variability among plankton communities was detected during summer. Overall, slight changes in plankton and fish composition resulted in the potential reorganization of the pelagic food web at the local scale. This work supports the utility of eDNA metaB in combination with hydrodynamic modelling to study marine biodiversity in the water column of coastal systems.

This article is part of the theme issue ‘Connected interactions: enriching food web research by spatial and social interactions’.

1. Introduction

Unlike on land, most primary and secondary producers are organisms smaller than a few millimetres in the ocean water column [1,2]. These floating communities belong to the plankton and tightly drive aquatic food webs [1]. Plankton have huge functional diversity, with autotrophs, heterotrophs, mixotrophs, herbivores, carnivores and detritivores coexisting at the microscale (up to 100 m) [3,4]. Such diversity displays unexpectedly long trophic pathways in a few cubic metres of seawater and feeds important

categories of fishes [5], like those playing as ‘keystones’ in marine food webs [6].

By definition, plankton drift with currents [7], and their communities show high spatiotemporal variability owing to the tight interplay between environmental factors and water transport [8,9]. On the one side, local physicochemical conditions select for plankton organisms with different physiology, biological cycles and feeding behaviours [9,10]; on the other side, ocean currents can reciprocally segregate or mix different water masses with distinct physicochemical properties which plankton can benefit from, or not [8,9,11]. This variability quickly scales up to fish communities [10,12], whose distribution in space and time is shaped by both evolutionary and ecological factors, from spawning timing and migratory abilities to salinity tolerance and food preferences [13–15].

Pelagic food webs, spanning from plankton to fish, can be, therefore, highly dynamic in space and time, especially at the regional scale (1–100 km) and close to the coast [16], where riverine inputs and intensified water flows can profoundly modify environmental conditions and the ecological state of the water column [17,18], with cascading effects on fish populations and pelagic food webs [12,19,20]. In this context, an important question is whether the variability of plankton–fish consortia, whose predator–prey relationships are a fundamental factor for fish recruitment, is higher in space or time in coastal systems, posing fundamental implications in the marine ecological study and management of ecosystem goods and services, and economic activities like fisheries and fish aquaculture [5,21].

This article is a proof-of-concept for the study of spatiotemporal changes in pelagic food webs in a coastal system at the regional scale (Campania region, Mediterranean Sea) and across different seasons (2020–2021). The study integrated the plankton–fish biodiversity inventory with the environmental DNA technique [22] and coastal connectivity (CC) analyses carried out with Lagrangian modelling to assess the probability that ocean currents transport plankton from one site to another over a given time interval [23]. Such an integrated approach allowed us to evaluate the degree of ecological and physical connectivity among communities and geographical sites and to assess the relative contribution of space and time in pelagic food web variability.

2. Material and methods

(a) Physical connectivity among coastal sites

To track estimated connectivity among nearshore sites we first performed numerical simulations using a regional ocean modelling system (ROMS) developed for the Tyrrhenian Sea (2 km resolution); then, the results of this first simulation were used as initial and boundary conditions for a finer grid model called Gulf of Naples Advanced Model (GNAM), covering the Campania coast with a 500 m resolution to obtain high-resolution output. GNAM is a free-surface, terrain-following, primitive equations ocean model widely used for a broad range of applications [24] and recently validated for the Gulf of Naples (GoN) area using a multiannual comparison with coastal high-frequency radar data and hydrological measurements [25]. We then used the ROMS velocity fields to run a Lagrangian transport package of virtual passive particles released along coastal areas, following velocity fields and constrained to fixed release depths (1 m).

CC is defined as the percentage of numerical particles, representing small water volumes and the plankton therein, leaving a source site (i) and arriving at a destination site (j) over a time interval t . Given n different coastal areas, an $n \times n$ connectivity matrix was evaluated for each given time scale, where the (i,j) element was the fraction of the particles from source area (i) to destination area (j), in the released time (t). In this study, we released particles along the Campania region coast (figure 1a) every 5 days for 5 years (2013–2017, around 250,000 particles per year), and tracked them for 96 days (for IDs, the number of release areas and the seasonal connectivity, see electronic supplementary material, table S1 and figure S1). Finally, a connectivity network was produced by summing the particle fraction of the areas of the sampling sites. Connectivity networks were visualized with Gephi v. 0.10 [26].

(b) Seawater sampling

We sampled environmental DNA (eDNA) onboard the *R/V Vettoria* between January 2020 and September 2021 in different sites along the coast of the Campania region (Southern Tyrrhenian Sea, Italy; figure 1a). Sampling in the GoN occurred at an approximately monthly scale; sampling at the plumes of three rivers in the GoN, Gulf of Gaeta (GoG) and Gulf of Salerno (GoS) occurred during summer (see electronic supplementary material, table S2). In the GoN, we sampled the long-term ecological research site MareChiara (DEIMS id: <https://deims.org/0b87459a-da3c-45af-a3e1-cb1508519411>) (40°48′ N, 14°15′ E) [27], the Sarno River mouth (40°43′ N, 14°27′ E), and an offshore site localized above a canyon, i.e. the Dohrn Canyon (40°36′ N, 14°08′ E). Other stations were at the Volturno River (40°58′ N, 13°50′ E) and Sele River (40°28′ N, 14°55′ E) mouths, in the GoG and GoS, respectively.

At each sampling site, we collected surface seawater using Niskin bottles, we filtered 0.5–2 L of seawater on nitrocellulose filters (porosity 0.45 μm , diameter 47 mm, GVS North America, two replicates per each sample) that were flash-frozen in liquid nitrogen and then preserved at -80°C until further analyses. A SeaBird 911 Plus multi-parametric probe provided temperature, salinity, density, conductivity, dissolved oxygen, fluorescence and turbidity data.

(c) Metabarcoding analyses

We extracted DNA using the E.Z.N.A. Mollusc DNA kit (Omega Bio-Tech) following the manufacturer’s instructions. Metabarcoding libraries were prepared using the primers Euk1391F and EukBr [28] for the V9-18S region and 12S MiFish_U forward

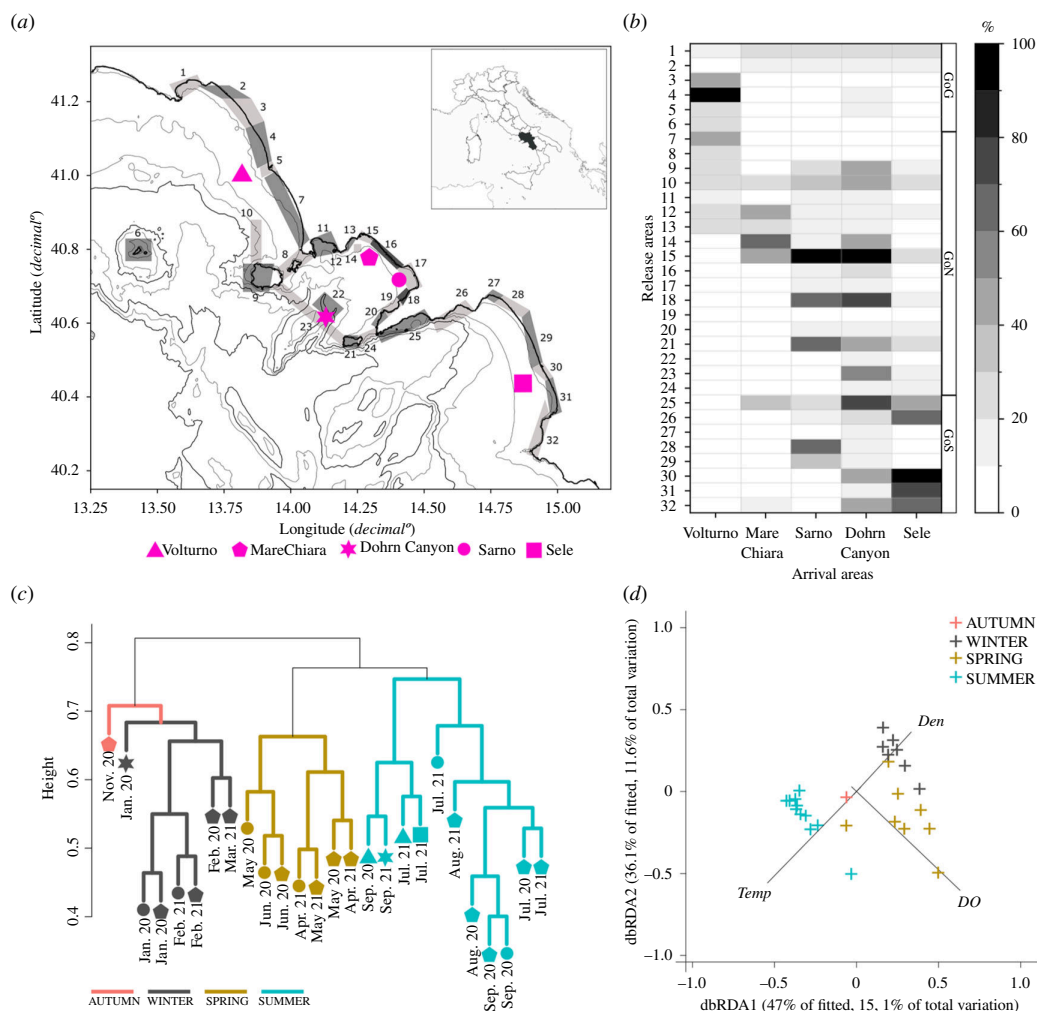


Figure 1. (a) Sampling map where numbers correspond to particle release areas (defined by the contiguous rectangles) along the Campanian coast; geometric shapes represent sampling stations. (b) Annual physical connectivity matrix showing particle migration rates, from release to arrival areas, as percentages (%). Cluster dendrogram (c) and dbRDA plot (d) based on Hellinger transformation of the reads count for V9-18S eDNA. (d) dbRDA plot fitted to significant predictor variables determined by BEST selection distLM (Temp, Temperature; DO, Dissolved Oxygen; Den, Density).

(fw) and reverse (rv) [29], respectively, and sequenced by Genomix4Life (Illumina MiSeq; <https://www.genomix4life.com/it/>) following published protocols [30,31]. The V9-18S region was chosen for its capability to detect most of the planktonic taxa, as already done in the GoN [32] and in other marine systems [33,34]. On the other hand, the 12S region was used only to selectively detect fish presence, since the number of reads referring to each amplicon sequence variant (ASV) did not allow for an estimate of the relative abundance of fish taxa.

Illumina paired-end V9-18S raw reads (FASTQ format) were pre-processed to generate ASVs in RStudio [35] using the dada2 pipeline [36]. Primer sequences were removed, and fw and rv reads were trimmed based on the quality score (the first 150 bases of each fw and rv reads were kept; the maximum number of 'expected errors' allowed in a read = 2; max number of ambiguities = 0). Filtered reads were used to train the error model from the data using a machine-learning approach. Fw and rv reads were then denoised to generate ASVs by applying the trained error model and using the option 'trimOverhang = TRUE' to account for the fact that the sequenced amplicon was smaller than the read size. Finally, fw and rv reads were merged and checked for chimeras. 12S ASVs were also generated with the dada2 R library; adapter trimming and preliminary filtering were instead performed using cutadapt [37] with the 'linked adapter' option in paired-end mode, allowing 20% mismatch, truncating 3' bases when quality was <15 and discarding untrimmed reads. All reads with ambiguities were then removed in dada2 before read error estimation and denoising; denoised reads were then merged into contigs, allowing a maximum of nine mismatches, and finally checked for chimeras.

To account for differences in the number of V9-18S region ASVs across samples, data were normalized at the median value of reads across samples ($n = 91,446$) using the function 'rrarefy' of the vegan R package [38]. Taxonomy was assigned to ASVs using a consensus taxonomy approach through the Python script 'taxonomy_assignment_BLAST.py' (https://github.com/Joseph7e/Assign-Taxonomy-with-BLAST/blob/master/taxonomy_assignment_BLAST.py) from five BLAST hits, using a minimum coverage of 70% and assigning taxa to species if the percentage of identity was $\geq 99\%$, to genus if $< 99\%$ and $\geq 95\%$, or to any other taxonomic categories if higher than such thresholds. The script was run twice, the first time against the SSU eukaryotic rRNA database of NCBI (https://ftp.ncbi.nlm.nih.gov/blast/db/SSU_eukaryote_rRNA.tar.gz, last modified 7 December 2022) and the second time against the PR2 database v4.14.0 (<https://github.com/pr2database/pr2database/releases>).

(d) Ecological data analysis

We performed statistical analyses using data without replicates and, where present, relative abundance was averaged, as done in similar studies (e.g. [33]), to a total of 26 eDNA samples. Environmental and biological (V9-18S eDNA) data were analysed separately (see electronic supplementary material, tables S2 and S3); the similarity matrices for environmental data were based on Euclidean distances of normalized data, while for biological data, we employed Hellinger transformation to reduce the impact of highly abundant taxa [39]. As the first exploratory analysis of beta-diversity, we conducted cluster analysis on V9-18S Hellinger-transformer data in the *vegan* package (*'decostand'*, *'vegdist'* and *'hclust'* functions) [40]. To test for the presence of seasonal differences between samples, we performed a two-way permutational multivariate analysis of variance (PERMANOVA, $p < 0.05$) with the fixed factor 'season' (three levels: winter, spring, summer), followed by a PAIRWISE test for significant terms.

We examined relationships between biological and environmental variables using the distance-based linear models (DistLM) routine that models linear relationships between dissimilarity matrices of biological data and predictor environmental variable(s) [41]. This routine allows fitting one or more environmental predictors to one or more biological variables. Among the model-building options, we selected the 'Best' procedure for the variables selection and 'An Information Criterion' ('AIC'; [42]) criterion for model comparisons. The criterion comes from the likelihood theory and smaller AIC values indicate a better model. Before the DistLM analysis, we used the Draftsman plot to reduce the effect of redundant variables and examine the correlation among environmental parameters before the analyses [43]. Conductivity, the only redundant variable showing >90% correlation, was excluded from the analyses. Statistical significance (PERMANOVA, $p < 0.05$) of the DistLM routine was assessed by permutation tests where each set of samples was randomly permuted 9999 times [44]. Analyses and plots were performed using the software PRIMER v.6.1.11 [44] and RStudio v.4.3.2 [35].

To investigate pelagic food webs, we identified potential trophic relationships between plankton and small pelagic fish detected, respectively, by V9-18S data and 12S on each site in different seasons. Therefore, plankton ASVs were summed and aggregated into specific functional groups (FGs) based on their taxonomic, dimensional [45,46] and physiological similarities [3]. The use of FGs is important as it increases the representation in food web models of ecological roles played by planktonic organisms within marine ecosystems, reducing complexity and functional redundancy [19,45]. Based on this rationale, we identified in our dataset seven FGs: autotrophic protists, heterotrophic/mixotrophic protists, crustaceans, gelatinous filter feeders, jellyfish, arrow worms and terrestrial organic matter (see electronic supplementary material, tables S3 and S4).

Planktonic FGs and small pelagic fish taxa were represented in conceptual food webs considering, respectively, their relative abundance (based on reads count) and presence-absence. Putative trophic interactions among FGs and fish were obtained from the literature and GloBi [47] (see electronic supplementary material, table S5). We represented conceptual pelagic food webs using the software Gephi, v. 0.10 [26].

3. Results

Lagrangian particle simulations showed a higher annual average connectivity among sampling stations in the GoN. Within these latter stations, the MareChiara site was largely connected to the Sarno River mouth, while the Volturno and Sele River mouths were less connected (figure 1b). However, model results showed that all sites were connected to different extents, thus allowing us to intercompare biological samples and their taxonomic composition studied with eDNA metabarcoding (eDNA metaB) and interpret spatial differences in light of connectivity among sites.

We annotated 4,344 planktonic and 13 fish ASV/taxa (seven pelagic and six demersal fish ASV) using the V9-18S and 12S regions, respectively (for more details about annotation results, see electronic supplementary material, tables S3 and S6). V9-18S samples showed seasonal partitioning, with three groups of samples, the first including all winter samples and one autumn, and the second and third groups, which differed more from the first group, including the spring and summer samples, respectively (figure 1c).

The PERMANOVA test found significant differences in biological data among seasons ($p < 0.05$). DistLM analysis showed that environmental parameters drove biological data partitioning, with temperature, density and dissolved oxygen explaining 26.7% of the total variance (figure 1d). The summer group was the largest one and included two subgroups, one including samples from the inner GoN (MareChiara and Sarno stations), and one including samples from the outer GoN (Dohrn Canyon) and other gulfs' sites. The latter observation indicates that spatial partitioning occurred, though to a lower extent than the seasonal one.

To map compositional differences in time and space, we combined plankton FGs and small pelagic fish presence and derived conceptual pelagic food webs (figure 2). MareChiara and Sarno stations had the best spatiotemporal coverage and strong connectivity all over the year (figure 1b), allowing seasonal comparisons (figure 2). At both sites, heterotrophic/mixotrophic protists were predominant and more abundant in winter and spring than in summer. Autotrophic protists were higher in summer at both sites. Crustaceans, which predate the previous groups, were third in rank and found at MareChiara during summer and winter but virtually absent during spring; however, during the latter season, crustaceans were detectable at Sarno. Other FGs were weakly detected overall.

Concerning small pelagic fish communities, we also observed signals of spatiotemporal differentiation at MareChiara and Sarno. The small pelagic *S. aurita* occurred in all the seasons analysed at the MareChiara and Sarno sites, while *E. encrasicolus* was always present but not at Sarno in spring. Winter was the richest season at the MareChiara and Sarno sites, which differed for the presence of *S. pilchardus* and *C. auratus* (present only at the MareChiara site).

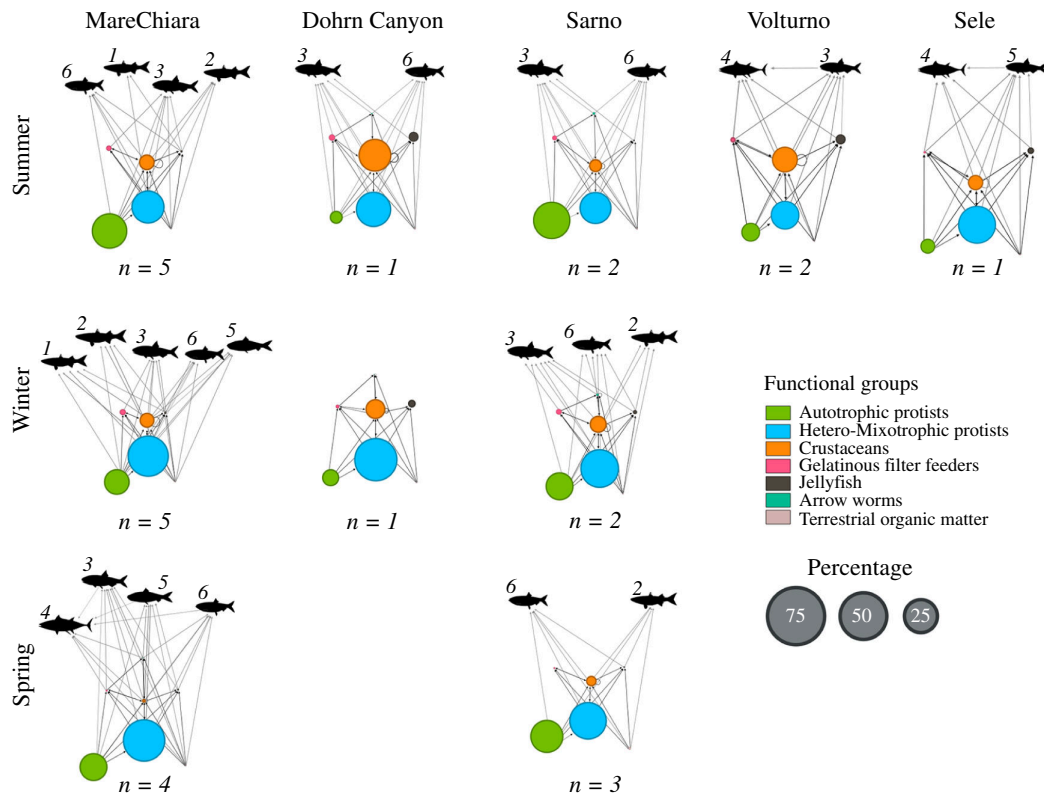


Figure 2. Conceptual pelagic food webs at the sampling sites in the summer, winter and spring seasons (samples collected during autumn, i.e. Nov_20 at the MareChiara station, were grouped into the winter samples). Edges represent putative trophic interactions between plankton FGs and small pelagic fish. Node size is proportional to the relative percentage of reads for V9-18S eDNA, while the information obtained from 12S (fish) was used to indicate the presence of taxa. Fish IDs: 1, *Chelon auratus*; 2, *C. labrosus*; 3, *Engraulis encrasicolus*; 4, *Euthynnus alletteratus*; 5, *Sardina pilchardus*; 6, *Sardinella aurita*. n is the number of samples used to calculate the percentage of total V9-18S reads.

Dohrn Canyon samples allowed us to describe only winter and summer communities, which were markedly different from those found in the inner GoN. About plankton, autotrophic protists were scanty if compared with MareChiara and Sarno, while heterotrophic/mixotrophic protists showed similar values. Crustaceans were in higher percentage during summer and showed almost double the relative abundance of the inner GoN. The other FGs were less represented, except for jellyfish, which showed a higher relative abundance than in the inner GoN. Finally, the Dohrn Canyon and Sarno sites showed the same summer small pelagic fish community.

We also explored the spatial differences among summer communities from the GoN and the other gulfs, i.e. studied at the Volturmo and Sele stations. The Volturmo plankton community was similar to that present at the Dohrn Canyon but showed a higher relative number of autotrophic protists. At Sele site, heterotrophic/mixotrophic protists were the most relevant, reaching the highest relative abundance among all summer communities. Still, autotrophic protists showed a lower amount than in other coastal sites during summer. Concerning planktonic animals, Volturmo was similar to the Dohrn Canyon (higher crustaceans and jellyfish than in other stations), while Sele was more similar to inner GoN stations (lower crustaceans and jellyfish). Concerning small pelagic fish, *E. alletteratus* and *E. encrasicolus* were present at both Volturmo and Sele, but this taxa combination was not found in any other site during summer.

4. Discussion

The advance in the eDNA metaB analysis in marine ecology has profoundly increased the amount of information available on marine biodiversity. The use of eDNA presents many advantages: for instance, it is less invasive and more cost-effective than traditional surveys [48,49]. So far the eDNA technique has been mainly applied to temporal or spatial studies with only a few investigations comparing the spatiotemporal changes of marine communities at the regional scale (e.g. [50,51]).

Our study shows the importance of integrating eDNA data and connectivity analysis to understand the dynamics of coastal planktonic communities, which are carried by physical dynamics and may be moved to different sites, influencing the higher trophic levels. Our results confirmed the plankton seasonality previously described at the MareChiara station (e.g. [32,52]), but extended this observation at the whole regional scale, with the planktonic communities in the Campania coast that differed more by seasonal than by spatial dynamics. This observation is in line with a recent scientific reference about plankton we can invoke as a comparison for the study area, i.e. the observation that the genetic fingerprint of populations of the diatom species *Pseudo-nitzschia multistriata* collected in the different Campania gulfs was spatially very similar, but far different over time [53].

Environmental factors determined these seasonal differences. Summer and winter communities detected with V9-18S eDNA metaB were mainly influenced by temperature and density, respectively, which remark the water column stratification cycles affecting nutrient availability in the photic zone influencing the metabolism, growth, reproduction, development, distribution

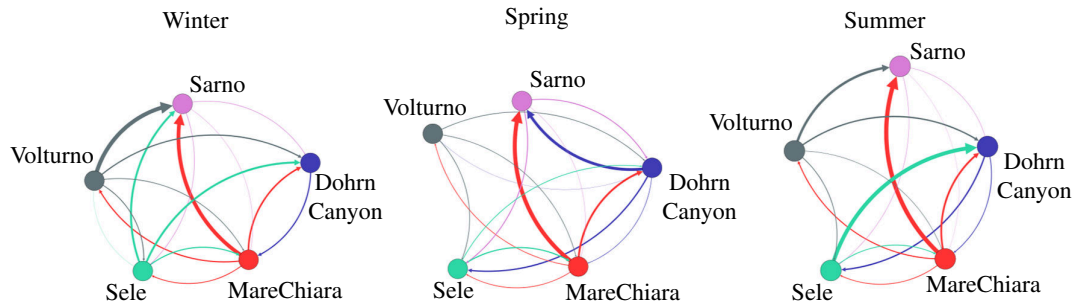


Figure 3. Connectivity network. Network edges represent connections between sites. Edges directed from release to the arrival site represent the particle release and are colour-coded by the release site. Edge width is proportional to the sum of the values of particle migration rate, grouped by the sampling site's area of interest.

and food availability of marine organisms [54,55]. The higher presence of phototrophs in summer and of heterotrophic/mixotrophic protists in winter we observed in the GoN matched that from a longer metabarcoding time series from the MareChiara site [32]. Spring communities are also influenced by dissolved oxygen (DO), which is regulated by both biotic (e.g. production and consumption), and abiotic factors (e.g. pH, temperature, salinity and hydrodynamic processes) [56,57]. DO concentrations can lead to changes in planktonic community structure and trophic transfer [58,59], with crustacean zooplankton peaking in hypoxic waters where they seek refuge from predation by fish, which instead exhibit lower abundance at lower oxygen levels [60,61].

Most of the spatial homogeneity that we detected could be the result of high connectivity at the regional scale (figure 3). For instance, the strong connection that between the Volturno and GoN areas matches previous modelling studies' results [62,63].

However, spatial differentiation can also emerge from the multiple stimuli that local environmental conditions may exert on the pelagic communities, which are highly dynamic entities. The tangled action of species immigration and local selection can affect spatial differentiation among communities and their food webs' functioning during the same season, like summer.

As a general pattern, phototroph-driven food webs and higher fish diversity occurred at eutrophic stations closer to the coast undergoing stronger inputs from land runoff and showing higher retention times, like in the inner GoN (MareChiara and Sarno), suggesting a more direct flux of matter from primary producers to higher consumers. Conversely, food webs at offshore stations (Dohrn Canyon), or those more exposed to the action of open sea currents (Volturno and Sele), showed a lower dependence on strictly phototrophic plankton, higher prevalence of heterotrophic/mixotrophic protists adapted to oligotrophic conditions, and less diverse small pelagic fish communities, suggesting a more dissipative microbial loop-based food web with longer trophic chains [64]. Oligotrophic sites also included more jellyfish, which compete with planktivorous fish [65], probably owing to upwelling transporting specimens from deeper waters [66].

Overall, we observed that slightly different plankton assemblages co-occurred with the different small pelagic fishes (figure 2) showing mainly a planktivorous diet composed of copepods, and to a lesser extent other crustaceans, molluscs, pelagic tunicates and other fishes [67,68]. The assembly of small pelagic fishes can be influenced by several interplaying drivers, such as food quality [13,14], biological (e.g. spawning timing) and environmental (e.g. salinity tolerance) factors [15,69].

Concerning biological factors, small pelagic fishes show different spawning times, which may have affected the occurrence of fish DNA in our study. Sardine (*S. pilchardus*, #5 in figure 2) spawns during autumn–winter in a mixed water column with salinities around 37–38 psu [68,69], which corresponds with the environmental conditions that we retrieved. Other species have their optimum reproduction and spawning during spring and summer when the water column is stratified and warmer, as in the case of skipjack tuna (*E. alletteratus*; #4) (23–27.5°C) and anchovy (*E. encrasicolus*; #3) (13–25.5°C) [70], which can spawn in a wide range of salinity (36.7–37.9 and 29.1–38.2 psu, respectively) [70,71]. Sardinella (*S. aurita*; #10) also spawns at temperatures above 23°C, from July to October in agreement with its tropical origin [69].

Regarding environmental factors, salinity tolerance can also drive fish assemblages. For instance, we found *C. auratus* (#1) only at MareChiara station, and *C. labrosus* (#2) also at the River Sarno mouth. This is in agreement with observations of *C. auratus* having mainly a pelagic behaviour, while *C. labrosus* is euryhaline and frequently found in estuaries [72]. The genus *Chelon* includes generalist planktivorous–detritivorous fish [73,74] often found around sea bream and sea bass farms [72,75] and polluted environments [76].

However, we must consider that the small pelagic fish species found in our survey may not reflect the entire diversity present in the study area. Indeed, fishes are characterized by a scattered spatial distribution [70]; water sampling can fail to catch all the taxa retrieved from traditional approaches, such as visual census and fishing nets [49,50]; eDNA shows operational limits like primer specificity [50], and it cannot provide information on the true abundances of organisms [47,48].

5. Conclusion

The application of eDNA metabarcoding analysis in marine ecology has expanded our understanding of the spatiotemporal dynamics of marine biodiversity at the regional scale. By applying an integrative approach, this study highlighted the importance of combining eDNA data with connectivity analyses to reveal the complex dynamics of coastal planktonic communities, which are influenced by physical processes and can spread between different sites affecting, in this way, the higher trophic levels, like fishes. In this respect, our study highlights the need to further investigate the intricate interactions that regulate

coastal marine communities and their underlying ecological processes, by intensifying the fish eDNA sampling effort at the spatial level and integrating these observations with traditional approaches like fishery surveys, which could provide more precise information on the distribution of fish in the water column. Such knowledge is critical for the effective conservation and management of marine ecosystems in the face of environmental change.

Ethics. This work did not require ethical approval from a human subject or animal welfare committee.

Data accessibility. The datasets supporting this article have been uploaded as part of the electronic supplementary material [77]. Raw V9-18S and 12S metabarcoding data are available in the Sequence Read Archive (SRA) under BioProject PRJNA1023387.

Declaration of AI use. We have not used AI-assisted technologies in creating this article.

Authors' contributions. D.B.: data curation, formal analysis, investigation, software, visualization, writing—original draft, writing—review and editing; L.R.: data curation, formal analysis, investigation, software, visualization, writing—original draft, writing—review and editing; V.D.T.: investigation; D.D.L.: data curation, investigation, methodology, visualization, writing—review and editing; G.D.G.: investigation; G.Z.: data curation, investigation, methodology, software; F.K.: data curation, investigation, methodology, visualization; V.B.: data curation, investigation, methodology; F.C.: investigation, writing—review and editing; F.C.: data curation, funding acquisition, resources; P.D.L.: methodology, resources; D.I.: funding acquisition, methodology, resources; F.M.: investigation, methodology, resources; S.S.: data curation, investigation, methodology, visualization; P.V.: funding acquisition, supervision; D.C.: conceptualization, methodology, supervision, writing—review and editing; D.D'A.: conceptualization, funding acquisition, methodology, supervision, writing—review and editing, project administration.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare we have no competing interests.

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References

1. Worden AZ, Follows MJ, Giovannoni SJ, Wilken S, Zimmerman AE, Keeling PJ. 2015 Environmental science. Rethinking the marine carbon cycle: factoring in the multifarious lifestyles of microbes. *Science* **347**, 1257594. (doi:10.1126/science.1257594)
2. Field CB, Behrenfeld MJ, Randerson JT, Falkowski P. 1998 Primary production of the biosphere: integrating terrestrial and oceanic components. *Science* **281**, 237–240. (doi:10.1126/science.281.5374.237)
3. Mitra A *et al.* 2023 The Mixoplankton Database (MDB): diversity of photo-phago-trophic plankton in form, function, and distribution across the global ocean. *J. Eukaryot. Microbiol.* **70**, e12972. (doi:10.1111/jeu.12972)
4. Benedetti F, Gasparini S, Ayata SD. 2016 Identifying copepod functional groups from species functional traits. *J. Plankton Res.* **38**, 159–166. (doi:10.1093/plankt/fbv096)
5. Lomartire S, Marques JC, Gonçalves AMM. 2021 The key role of zooplankton in ecosystem services: a perspective of interaction between zooplankton and fish recruitment. *Ecol. Indic.* **129**, 107867. (doi:10.1016/j.ecolind.2021.107867)
6. Coll M, Libralato S. 2012 Contributions of food web modelling to the ecosystem approach to marine resource management in the Mediterranean Sea. *Fish Fish.* **13**, 60–88. (doi:10.1111/j.1467-2979.2011.00420.x)
7. Hensen V. 1887 Kapitel 1: Über die Bestimmung des Plankton's oder des im Meere treibenden materials an Pflanzen und Thieren. *Jahresber. Comm. Zur Wiss. Unters. Dtsch. Meere Kiel Für Jahre* **12**, 1–107.
8. Jagadeesan L, Jyothibabu R, Anjusha A, Mohan AP, Madhu NV, Muraleedharan KR, Sudheesh K. 2013 Ocean currents structuring the mesozooplankton in the Gulf of Mannar and the Palk Bay, southeast coast of India. *Prog. Oceanogr.* **110**, 27–48. (doi:10.1016/j.pocan.2012.12.002)

9. Bialonski S, Caron DA, Schloen J, Feudel U, Kantz H, Moorthi SD. 2016 Phytoplankton dynamics in the Southern California Bight indicate a complex mixture of transport and biology. *J. Plankton Res.* **38**, 1077–1091. (doi:10.1093/plankt/fbv122)
10. Wei Y, Cui Z, Shi Y, Shan X, Chen B, Qu K, Xin Q, Jiang T, Chen J. 2023 Contrasting currents drive geographic variability in the biomass of Pacific saury (*Cololabis saira*), zooplankton, and phytoplankton in the northwestern Pacific. *Prog. Oceanogr.* **217**, 103099. (doi:10.1016/j.pocean.2023.103099)
11. Jönsson BF, Watson JR. 2016 The timescales of global surface-ocean connectivity. *Nat. Commun.* **7**, 11239. (doi:10.1038/ncomms11239)
12. Cushing DH. 1995 The long-term relationship between zooplankton and fish: IV. spatial/temporal variability and prediction. *ICES J. Mar. Sci.* **52**, 611–626. (doi:10.1016/1054-3139(95)80076-X)
13. Whitfield AK. 1988 The fish community of the Swartvlei estuary and the influence of food availability on resource utilization. *Estuaries* **11**, 160–170. (doi:10.2307/1351968)
14. Tableau A, Brind'Amour A, Woillez M, Le Bris H. 2016 Influence of food availability on the spatial distribution of juvenile fish within soft sediment nursery habitats. *J. Sea Res.* **111**, 76–87. (doi:10.1016/j.seares.2015.12.004)
15. Akimova A, Núñez-Riboni I, Kempf A, Taylor MH. 2016 Spatially-resolved influence of temperature and salinity on stock and recruitment variability of commercially important fishes in the North Sea. *PLoS One* **11**, e0161917. (doi:10.1371/journal.pone.0161917)
16. Hernández-Carrasco I, Orfila A, Rossi V, Garçon V. 2018 Effect of small scale transport processes on phytoplankton distribution in coastal seas. *Sci. Rep.* **8**, 8613. (doi:10.1038/s41598-018-26857-9)
17. Stenseth NC, Llope M, Anadón R, Ciannelli L, Chan KS, Hjermann DØ, Bagøien E, Ottersen G. 2006 Seasonal plankton dynamics along a cross-shelf gradient. *Proc. R. Soc. B* **273**, 2831–2838. (doi:10.1098/rspb.2006.3658)
18. DiBattista JD, Reimer JD, Stat M, Masucci GD, Biondi P, De Brauwier M, Wilkinson SP, Chariton AA, Bunce M. 2020 Environmental DNA can act as a biodiversity barometer of anthropogenic pressures in coastal ecosystems. *Sci. Rep.* **10**, 8365. (doi:10.1038/s41598-020-64858-9)
19. Russo L, Casella V, Marabotti A, Jordán F, Congestri R, D'Alelio D. 2022 Trophic hierarchy in a marine community revealed by network analysis on co-occurrence data. *Food Webs* **32**, e00246. (doi:10.1016/j.fooweb.2022.e00246)
20. Vassallo P, Bellardini D, Castellano M, Daputo G, Povero P. 2022 Structure and functionality of the mesozooplankton community in a coastal marine environment: Portofino marine protected area (Liguria). *Diversity* **14**, 19. (doi:10.3390/d14010019)
21. Nissar S, Bakhtiyar Y, Arafat MY, Andrabi S, Bhat AA, Yousuf T. 2023 A review of the ecosystem services provided by the marine forage fish. *Hydrobiologia* **850**, 2871–2902. (doi:10.1007/s10750-022-05033-1)
22. Taberlet P, Coissac E, Pompanon F, Brochmann C, Willerslev E. 2012 Towards next-generation biodiversity assessment using DNA metabarcoding. *Mol. Ecol.* **21**, 2045–2050. (doi:10.1111/j.1365-294X.2012.05470.x)
23. Mitarai S, Siegel DA, Watson JR, Dong C, McWilliams JC. 2009 Quantifying connectivity in the coastal ocean with application to the Southern California Bight. *J. Geophys. Res.* **114**, C10026. (doi:10.1029/2008JC005166)
24. Haidvogel DB *et al.* 2008 Ocean forecasting in terrain-following coordinates: formulation and skill assessment of the Regional Ocean Modeling System. *J. Comput. Phys.* **227**, 3595–3624. (doi:10.1016/j.jcp.2007.06.016)
25. Kokoszka F, Saviano S, Botte V, Ludicone D, Zambianchi E, Cianelli D. 2022 Gulf of Naples advanced model (GNAM): a multiannual comparison with coastal hf radar data and hydrological measurements in a coastal Tyrrhenian basin. *J. Mar. Sci. Eng.* **10**, 1044. (doi:10.3390/jmse10081044)
26. Bastian M, Heymann S, Jacomy M. 2009 Gephi: an open source software for exploring and manipulating networks. *ICWSM* **3**, 361–362. (doi:10.1609/icwsm.v3i1.13937)
27. Zingone A, Tortora C, D'Alelio D, Margiotta F, Sarno D. 2023 Assembly rules vary seasonally in stable phytoplankton associations of the Gulf of Naples (Mediterranean Sea). *Mar. Ecol.* **44**, e12730. (doi:10.1111/maec.12730)
28. Amaral-Zettler LA, McCliment EA, Ducklow HW, Huse SM. 2009 A method for studying protistan diversity using massively parallel sequencing of V9 hypervariable regions of small-subunit ribosomal RNA genes. *PLoS One* **4**, e6372. (doi:10.1371/journal.pone.0006372)
29. Miya M *et al.* 2015 MiFish, a set of universal PCR primers for metabarcoding environmental DNA from fishes: detection of more than 230 subtropical marine species. *R. Soc. Open Sci.* **2**, 150088. (doi:10.1098/rsos.150088)
30. Closek C *et al.* 2018 Environmental DNA (Edna) 18S Metabarcoding Illumina Miseq NGS PCR protocol V2. protocols.io. (doi:10.17504/protocols.io.n2vdge6)
31. Closek C *et al.* 2018 Environmental DNA (Edna) 12S Metabarcoding Illumina Miseq NGS PCR protocol V1. protocols.io. (doi:10.17504/protocols.io.m3bc8in)
32. Piredda R, Tomasino MP, D'Erchia AM, Manzari C, Pesole G, Montresor M, Kooistra WHCF, Sarno D, Zingone A. 2017 Diversity and temporal patterns of planktonic protist assemblages at a Mediterranean Long Term Ecological Research site. *FEMS Microbiol. Ecol.* **93**, fiw200. (doi:10.1093/femsec/fiw200)
33. Djurhuus A *et al.* 2020 Environmental DNA reveals seasonal shifts and potential interactions in a marine community. *Nat. Commun.* **11**, 254. (doi:10.1038/s41467-019-14105-1)
34. de Vargas C *et al.* 2015 Eukaryotic plankton diversity in the sunlit ocean. *Science* **348**, 1261605. (doi:10.1126/science.1261605)
35. RStudio Team. 2020 *Rstudio: integrated development for R. Rstudio*. See <http://www.rstudio.com/>.
36. Callahan BJ, McMurdie PJ, Rosen MJ, Han AW, Johnson AJA, Holmes SP. 2016 DADA2: high-resolution sample inference from Illumina amplicon data. *Nat. Methods* **13**, 581–583. (doi:10.1038/nmeth.3869)
37. Martin M. 2011 Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnet. J.* **17**, 10–12. (doi:10.14806/ej.17.1.200)
38. Oksanen J. 2022 Vegan: community Ecology package. See <http://vegan.r-forge.r-project.org/>.
39. Legendre P, Gallagher ED. 2001 Ecologically meaningful transformations for ordination of species data. *Oecologia* **129**, 271–280. (doi:10.1007/s004420100716)
40. McMurdie PJ, Holmes S. 2013 phyloseq: an R package for reproducible interactive analysis and graphics of microbiome census data. *PLoS One* **8**, e61217. (doi:10.1371/journal.pone.0061217)
41. McArdle BH, Anderson MJ. 2001 Fitting multivariate models to community data: a comment on distance-based redundancy analysis. *Ecology* **82**, 290–297. (doi:10.1890/0012-9658(2001)082[0290:FMTCDD]2.0.CO;2)
42. Akaike H. 1974 A new look at the statistical model identification. *IEEE Trans. Automat. Contr.* **19**, 716–723. (doi:10.1109/TAC.1974.1100705)
43. Clarke KR, Green RH. 1988 Statistical design and analysis for a 'biological effects' study. *Mar. Ecol. Prog. Ser.* **46**, 213–226. (doi:10.3354/meps046213)
44. Anderson M. 2008 *Permanova+ for primer: guide to software and statistical methods*. Plymouth, UK: Primer-E Ltd.
45. Boyce DG, Frank KT, Leggett WC. 2015 From mice to elephants: overturning the 'one size fits all' paradigm in marine plankton food chains. *Ecol. Lett.* **18**, 504–515. (doi:10.1111/ele.12434)
46. Ruggiero MA, Gordon DP, Orrell TM, Bailly N, Bourgoin T, Brusca RC, Cavalier-Smith T, Guiry MD, Kirk PM. 2015 A higher level classification of all living organisms. *PLoS One* **10**, e0119248. (doi:10.1371/journal.pone.0119248)

47. Poelen JH, Simons JD, Mungall CJ. 2014 Global biotic interactions: an open infrastructure to share and analyze species–interaction datasets. *Ecol. Inform.* **24**, 148–159. (doi:10.1016/j.ecoinf.2014.08.005)
48. Coble AA, Flinders CA, Homyack JA, Penaluna BE, Cronn RC, Weitemier K. 2019 eDNA as a tool for identifying freshwater species in sustainable forestry: a critical review and potential future applications. *Sci. Total Environ.* **649**, 1157–1170. (doi:10.1016/j.scitotenv.2018.08.370)
49. Hansen BK, Bekkevold D, Clausen LW, Nielsen EE. 2018 The sceptical optimist: challenges and perspectives for the application of environmental DNA in marine fisheries. *Fish Fish. (Oxf.)* **19**, 751–768. (doi:10.1111/faf.12286)
50. Berry TE *et al.* 2023 A 3-year plankton DNA metabarcoding survey reveals marine biodiversity patterns in Australian coastal waters. *Div. Dist.* **29**, 862–878. (doi:10.1111/ddi.13699)
51. Costalago D, Forster I, Nemcek N, Neville C, Perry RI, Young K, Hunt BPV. 2020 Seasonal and spatial dynamics of the planktonic trophic biomarkers in the Strait of Georgia (northeast Pacific) and implications for fish. *Sci. Rep.* **10**, 8517. (doi:10.1038/s41598-020-65557-1)
52. Di Capua I, Piredda R, Mazzocchi MG, Zingone A. 2021 Metazoan diversity and seasonality through eDNA metabarcoding at a Mediterranean long-term ecological research site. *ICES J. Mar. Sci.* **78**, 3303–3316. (doi:10.1093/icesjms/fsab059)
53. Ruggiero MV, D'Alelio D, Ferrante MI, Santoro M, Vitale L, Procaccini G, Montresor M. 2018 Clonal expansion behind a marine diatom bloom. *ISME J.* **12**, 463–472. (doi:10.1038/ismej.2017.181)
54. Staehr PA, Sand-Jensen K. 2006 Seasonal changes in temperature and nutrient control of photosynthesis, respiration and growth of natural phytoplankton communities. *Freshw. Biol.* **51**, 249–262. (doi:10.1111/j.1365-2427.2005.01490.x)
55. Zhao Q, Liu S, Niu X. 2020 Effect of water temperature on the dynamic behavior of phytoplankton–zooplankton model. *Appl. Math. Comput.* **378**, 125211. (doi:10.1016/j.amc.2020.125211)
56. Mandal S, Debnath M, Ray S, Ghosh PB, Roy M, Ray S. 2012 Dynamic modelling of dissolved oxygen in the creeks of Sagar Island, Hooghly–Matla estuarine system, West Bengal, India. *Appl. Math. Model.* **36**, 5952–5963. (doi:10.1016/j.apm.2011.10.013)
57. Banerjee A, Chakrabarty M, Rakshit N, Bhowmick AR, Ray S. 2019 Environmental factors as indicators of dissolved oxygen concentration and zooplankton abundance: deep learning versus traditional regression approach. *Ecol. Indic.* **100**, 99–117. (doi:10.1016/j.ecolind.2018.09.051)
58. Hull V, Mocenni C, Falcucci M, Marchettini N. 2000 A trophodynamic model for the lagoon of Fogliano (Italy) with ecological dependent modifying parameters. *Ecol. Modell.* **134**, 153–167. (doi:10.1016/S0304-3800(00)00358-6)
59. Hull V, Parrella L, Falcucci M. 2008 Modelling dissolved oxygen dynamics in coastal lagoons. *Ecol. Modell.* **211**, 468–480. (doi:10.1016/j.ecolmodel.2007.09.023)
60. Roman MR, Brandt SB, Houde ED, Pierson JJ. 2019 Interactive effects of hypoxia and temperature on coastal pelagic zooplankton and fish. *Front. Mar. Sci.* **6**, 00139. (doi:10.3389/fmars.2019.00139)
61. Weinstock JB, Vargas L, Collin R. 2022 Zooplankton abundance reflects oxygen concentration and dissolved organic matter in a seasonally hypoxic estuary. *J. Mar. Sci. Eng.* **10**, 427. (doi:10.3390/jmse10030427)
62. Iermano I, Moore AM, Zambianchi E. 2016 Impacts of a 4-dimensional variational data assimilation in a coastal ocean model of southern Tyrrhenian Sea. *J. Mar. Syst.* **154**, 157–171. (doi:10.1016/j.jmarsys.2015.09.006)
63. Ciannelli L, Cannavacciuolo A, Konstantinidis P, Mirasole A, Wong-Ala JATK, Guerra MT, D'Ambra I, Riginella E, Cianelli D. 2022 Ichthyoplankton assemblages and physical characteristics of two submarine canyons in the south central Tyrrhenian Sea. *Fish. Oceanogr.* **31**, 480–496. (doi:10.1111/fog.12596)
64. Azam F, Fenchel T, Field J, Gray J, Meyer-Reil L, Thingstad F. 1983 The ecological role of water-column microbes in the sea. *Mar. Ecol. Prog. Ser.* **10**, 257–263. (doi:10.3354/meps010257)
65. Brodeur RD, Suchman CL, Reese DC, Miller TW, Daly EA. 2008 Spatial overlap and trophic interactions between pelagic fish and large jellyfish in the northern California Current. *Mar. Biol.* **154**, 649–659. (doi:10.1007/s00227-008-0958-3)
66. Bouillon J, Medel MD, Pagès F, Gili JM, Boero F, Gravili C. 2004 Fauna of the Mediterranean Hydrozoa. *Sci. Mar.* **68**, 5–438. (doi:10.3989/scimar.2004.68s25)
67. Taieb AH, Sley A, Ghorbel M, Jarboui O. 2013 Feeding habits of *Sparus aurata* (Sparidae) from the Gulf of Gabes (central Mediterranean). *Cah. Biol. Mar.* **54**, 263–270.
68. Morote E, Olivar MP, Villate F, Uriarte I. 2010 A comparison of anchovy (*Engraulis encrasicolus*) and sardine (*Sardina pilchardus*) larvae feeding in the Northwest Mediterranean: influence of prey availability and ontogeny. *ICES J. Mar. Sci.* **67**, 897–908. (doi:10.1093/icesjms/fsp302)
69. Palomera I, Olivar MP, Salat J, Sabatés A, Coll M, García A, Morales-Nin B. 2007 Small pelagic fish in the NW Mediterranean Sea: an ecological review. *Prog. Oceanogr.* **74**, 377–396. (doi:10.1016/j.pocean.2007.04.012)
70. Palomera I. 1992 Spawning of anchovy *Engraulis encrasicolus* in the Northwestern Mediterranean relative to hydrographic features in the region. *Mar. Ecol. Prog. Ser.* **79**, 215–223.
71. Alemany F, Quintanilla L, Velez-Belchí P, García A, Cortés D, Rodríguez JM, Fernández de Puelles ML, González-Pola C, López-Jurado JL. 2010 Characterization of the spawning habitat of Atlantic bluefin tuna and related species in the Balearic Sea (western Mediterranean). *Prog. Oceanogr.* **86**, 21–38. (doi:10.1016/j.pocean.2010.04.014)
72. Arechavala-Lopez P, Uglem I, Sanchez-Jerez P, Fernandez-Jover D, Bayle-Sempere JT, Nilsen R. 2010 Movements of grey mullet *Liza aurata* and *Chelon labrosus* associated with coastal fish farms in the western Mediterranean Sea. *Aquacult. Environ. Interact.* **1**, 127–136. (doi:10.3354/aei00012)
73. Ali M, And SA. 2022 Feeding habits of golden grey mullet *Liza aurata* (Risso, 1810) in the Bitter Lakes, Suez Canal, Egypt. *J. Anim. Poult. Fish Prod.* **11**, 9–14. (doi:10.21608/japfp.2022.284087)
74. Wassef EA, El Masry MH, Mikhail FR. 2001 Growth enhancement and muscle structure of striped mullet, *Mugil cephalus* L., fingerlings by feeding algal meal-based diets. *Aquac. Res.* **32**, 315–322. (doi:10.1046/j.1355-557x.2001.00043.x)
75. Fernandez-Jover D, Sanchez-Jerez P, Bayle-Sempere JT, Valle C, Dempster T. 2008 Seasonal patterns and diets of wild fish assemblages associated with Mediterranean coastal fish farms. *ICES J. Mar. Sci.* **65**, 1153–1160. (doi:10.1093/icesjms/fsn091)
76. Bakhshalizadeh S, Liyafoyi AR, Fazio F, Mora-Medina R, Ayala-Soldado N. 2023 Health risk assessment of heavy metal concentration in muscle of *Chelon auratus* and *Chelon saliens* from the southern Caspian Sea. *Environ. Geochem. Health* **45**, 3377–3385. (doi:10.1007/s10653-022-01401-x)
77. Bellardini D, Russo L, Di Tuccio V, De Luca D, Del Gaizo G, Zampicini G. 2024 Supplementary material from: Spatiotemporal changes of Pelagic food webs investigated by environmental DNA Metabarcoding and Connectivity analysis. Figshare. (doi:10.6084/m9.figshare.c.7294420)